Wilder Modification of the Bielschowsky Stain for Reticulum

Catalog #: 26392-Series

Fixation:

10% Buffered Neutral Formalin (#15740) or Russell's Zenker (#64123-05).

Sections:

Cut paraffin at 6µm sections.

Staining Procedures:

- 1. Deparaffinize and hydrate slides to distilled water.
- 2. Oxidize in <u>Phosphomolybdic Acid, 10% (#26392-07</u>) for 1 minute. Rinse well in distilled water or sunning water to prevent the cells from holding yellow colors.
- 3. Sensitize in <u>Uranyl Nitrate, 1% (#26392-08</u>) for 1 minute or less. Rinse 10 20 seconds in distilled water.
- 4. Stain in fresh Ammonical Silver for one minute (changing solution frequently). The solution may be prepared as follows:
 - 1. Add concentrate <u>Ammonium Hydroxide</u> (<u>#26392-02</u>) to 5.0 ml of <u>Silver Nitrate, 10.2%</u> (<u>#26392-01</u>) drop by drop, until the forming precipitate is almost dissolved.
 - Add 5.0 ml of <u>Sodium Hydroxide, 3.1%</u> (#26392-03) and just dissolve the resultant precipitate by slowly adding conc. Ammonium Hydroxide dropwise. Bring volume up to 50 ml with distilled water.
 - 3. Use at once or filter and store in refrigerator in amber bottle.
- 5. Dip quickly in 95% alcohol and transfer to freshly prepared reducing solution. Prepare the reducing solution as follows:
 - 1. Mix 50.0 ml distilled water, 1.5ml <u>Uranyl Nitrate, 1%</u> (#26392-08) and 0.5 ml of <u>Formalin, 40% neutralized</u> (#26392-04).
 - 2. Prepare just before use and change frequently during the process.
- 6. Rinse in distilled water.
- 7. Tone in <u>Gold Chloride, 0.2%</u> (#26392-09) approx. 1 minute, until the yellow brown color leaves the sections and they turn lavender. Check the slides at this point individually with a microscope to prevent over toning; this could lead to red sections.
- 8. Rinse in distilled water and transfer to <u>Sodium Thiosulfate</u>, <u>5%</u> (<u>#26392-10</u>) approx. 1 minute to fix the color. Wash in tap water.
- Counter-stain in <u>Nuclear Fast Red (#26392-06</u>) if desired, rinsing well in distilled water. Alternatively <u>Harris Hematoxylin (#26392-05</u>) with either <u>Van Gieson (#26392-5A</u>) or phloxine can be used as a counterstain.
- 10. Dehydrate in 95% and 100% alcohol, 2 changes and clear in xylene, 2 changes.
- 11. Mount. If celloidin is used, clear in oil of origanium followed by 25% alcohol, usually mounted in balsam with Permount.

Stain Results:

Reticulum fiber (If beading is observed and/or dark background, replace the	Black
ammonium Hydroxide Silver Nitrate solution and the reducing solution	
Collagen	Rose
Other tissue elements	Red

References:

Wilder, H.C., Amer. J. Clin. Path., 11:817, 1935.Mallory, F.B. Pathological Technique, Phil., W.B. Saunders & Co, c. 1938AFIP Manual of Histological Staining Techniques, NY, McGraw-Hill Publ., c. 1968/3rd ed. P.92